

Quality control assessment of HIV-2 viral load quantification assays

Results from an international collaboration on HIV-2 infection, 2006

Florence Damond¹ and Antoine Bénard², on behalf of the ACHIEV_{2E} study group

1. Laboratoire de Virologie, Hôpital Bichat – Claude Bernard, Paris, France
2. INSERM U593, ISPÉD, Université Victor Segalen Bordeaux 2, Bordeaux, 33076 France

ACHIEV_{2E} study group: Belgium: Patrick GOUBAU and Jean RUELLE (ADS Reference Laboratory, UCL); France: Françoise BRUN-VEZINET, Pauline CAMPA, Florence DAMOND, Sophie MATHERON (Bichat – Claude Bernard Hospital, Paris), Antoine BENARD, Geneviève CHÉNE, and Audrey TASEI (INSERM U593, Bordeaux 2 University, Bordeaux); Gambia: Abraham ALABI and Sarah ROWLAND-JONES (Medical Research Council); Germany: Bernd KUPFER (Institute for Medical Microbiology, Immunology, and Parasitology, Bonn); Jorge ROCKSTRÖM and Carolina SCHWARZ-ZANDER (Department of Internal Medicine I, University of Bonn, Bonn); Netherlands: Frank DE WOLFF, Pieter REISS (National AIDS Therapy Evaluation Center, ASC, Amsterdam) and Maarten SCHIM VAN DER LOEFF (AMC-CPIC, Amsterdam and MRC, Gambia); Portugal: Ricardo CAMACHO, Peralva GOMES and Karim MANSURHO (Egas Moniz Hospital, Lisbon); Francisco ANTUNES and Emília VALADÃO (Clínica Universitária de Doenças Infecciosas, Lisbon); Spain: Berta RODRÍGUEZ and Victoria SORIANO (Department of Infectious Diseases, Hospital Carlos III, Madrid, Spain); Sweden: Jan ALBERT (Dept of Virology, Swedish Institute for Infectious Disease control, Solna); Switzerland: Jörg KUHL (St. Gallen HIV Cohort Study); United Kingdom: Jonathan TOSSELL (Centre for Infectious Health Protection Agency, London), Bridget FERNS and Jeremy GARDSON (Centre for Virology, Royal Free & University College London Medical School, London).

BACKGROUND

There is currently no gold standard for the quantification of plasma HIV-2-RNA. At the patient level, difficulties arise for the interpretation of results and may yield inappropriate clinical decision; at the population level, this may explain controversial evaluation of response to treatment among different cohorts.

We evaluated the validity of HIV-2 subtype A RNA quantification assays performed in reference laboratories located in 9 countries (Belgium, France, Gambia, Germany, Portugal, Spain, Sweden, Switzerland and UK) within the ACHIEV_{2E} study group.

MATERIAL AND METHODS

In a double blind experimental design, each of the nine participating laboratories quantified:

- 3 series of 10 aliquots of NIHZ HIV-2 subtype A supernatant counted by electronic microscopy (ABI technologies) previously diluted in HIV negative human plasma and stored at -80°C. Each series was diluted at a different theoretical viral load: 1.7, 2.3 and 3 log₁₀ copies/ml.

- 5 HIV-2 negative aliquots.

Laboratories used mainly real PCR assays, with primers and probes located on various regions of HIV-2 genome and different HIV-2 standards.

Accuracy and precision of assays were estimated for the 3 theoretical concentration levels. A quantification assay was defined as accurate if at least 9/10 of the quantifications were in the expected interval:

$$[(\text{theoretical viral load}) \times 3 - (\text{theoretical viral load}) \times 3]$$

Precision of assays was evaluated using the intralaboratory coherence coefficient (ILCC) and the coefficient of variation (CV) at each of the 3 theoretical concentration viral loads.

$$ILCC_{ij} = \frac{s_{ij} \sqrt{p_j}}{\sqrt{\sum_{ij} s_{ij}^2}}$$

where i = laboratory
 j = concentration level
 p = number of laboratory
 s = standard deviation
 s^2 = variance

A quantification heterogeneity exists when $ILCC > L$
(L, extracted from a reference table for p laboratories and n repetitions)

$$CV_{ij} = \frac{s_{ij}}{m_{ij}}$$

where s_{ij} = standard deviation of the laboratory i at the concentration level j
 m_{ij} = mean of the quantifications of the laboratory i at the concentration level j

RESULTS

Characteristics of each evaluated quantification assays are shown in table 1. No false positive result was reported.

Table 1. Characteristics of quantification assays assessed in the ACHIEV_{2E} Collaboration, 2006.

Laboratories	System	primers and probes localization	RNA extraction	Standard used	Threshold (log ₁₀ copies/ml)	subtyped	Primer supplier	Probe supplier
1	Light cycler 2.0	LTR region (Green detection)	Nucleic Acid Amplification	external, synthetic RNA (ROX mix)	1.7	A,B	Campenon	No probe used
2	Light cycler	gag gene	Magnopure	external, MN2 quantified by electronic microscopy	2.0	A,B,H	MWG, probe and applied	Prigo
3	In-house PCR (SLONA)	LTR	Roan	CSL23 + internal control	2.0	A,B	MWG	MWG
4	nested PCR	LTR gag, gag, gag, gag, gag	Qiagen, viral DNA	external/MNV-11971	2.7	no reference	TIB MolBio	TIB MolBio
5	In house QuRT-PCR	Env gene and gag region	amplifying Bioblocks	internal control	2.3	A,B	Invitrogen	Invitrogen
6	Nucleic Acid Amplifier v1.1	gag gene	Nucleic Acid Amplification	internal	2.3	A,B	Provided in the Nucleic Acid Amplifier v1.1 (BioMérieux)	
7	SeqViral Load Analyser v1.0		seqViral RT test from Codi	external, RT test, only relative quantification	2.6	A, S/Other		
8	Thaiflon	gag leader	HIV Monitor	external, RT test, only relative quantification	not yet evaluated	A,B	Microsynth	Microsynth
9	ABI Prism 7000	LTR	Qiagen viral RNA	CSL23 external & BMV internal	2.0	A,B	MWG	MWG

At the concentration level of 3 log₁₀ copies/ml, five laboratories reported accurate estimations (n² 4, 5, 6, 9 ; figure 1) and seven reported precise estimations (CV= 3 to 6%; ILCC= 0.5 to 0.9) (n¹ 1, 2, 3, 6, 7, 8, 9 ; table 2).

At the concentration level of 2.3 log₁₀ copies/ml, three laboratories reported accurate estimations (n² 6, 9 ; figure 2) and six reported precise estimations (CV= 5 to 8%; ILCC= 0.5 to 1.1) (n¹ 1, 2, 3, 7, 8, 9 ; table 3).

At the concentration level of 1.7 log₁₀ copies/ml, two laboratories reported accurate estimations (n² 2 and 9), but with lack of precision (CV=24% and 33%; ILCC=1.3 and 1.5 respectively), and three reported precise estimations (CV= 5% to 11%; ILCC= 0.4 to 0.8) (n¹ 1, 3, 8).

CONCLUSIONS

HIV-2 subtype A viral load quantifications varied considerably between laboratories. If the NIHZ HIV-2 supernatant counted by electronic microscopy is regarded as a standard, only 2 laboratories reported accurate and precise estimations at the viral loads of both 3 and 2.3 log₁₀ copies/ml. The ACHIEV_{2E} network is working towards standardization of quantification assays to better interpret the response to treatment in HIV-2 infected patients and implement international clinical trials in the future.

Accuracy of quantification assays

Figure 1. Theoretical viral load = 3.0 log₁₀ cp/ml

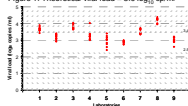


Figure 2. Theoretical viral load = 2.3 log₁₀ cp/ml

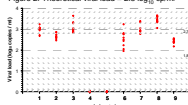
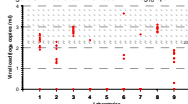


Figure 3. Theoretical viral load = 1.7 log₁₀ cp/ml



Precision of quantification assays

Table 2. Theoretical viral load = 3.0 log₁₀ cp/ml

Laboratories	Coefficient of variation (%)	Intra-laboratory precision (s.d./2.3)	Inter-laboratory precision (s.d./2.3)
1	4	0.71	NA
2	4	0.60	NA
3	4	0.76	NA
4	5	1.06	NA
5	4	0.70	0.62
6	10	1.03	0.43
7	3	0.60	NA
8	4	0.79	NA
9	6	0.69	NA

Table 3. Theoretical viral load = 2.3 log₁₀ cp/ml

Laboratories	Coefficient of variation (%)	Intra-laboratory precision (s.d./2.3)	Inter-laboratory precision (s.d./2.3)
1	5	0.71	NA
2	6	0.60	NA
3	6	1.06	NA
4	6	1.06	NA
5	5	0.70	0.62
6	16	1.03	0.43
7	11	0.60	0.43
8	6	0.79	NA
9	6	0.69	NA

Table 4. Theoretical viral load = 1.7 log₁₀ cp/ml

Laboratories	Coefficient of variation (%)	Intra-laboratory precision (s.d./1.7)	Inter-laboratory precision (s.d./1.7)
1	5	0.71	NA
2	24	1.06	NA
3	6	0.60	NA
4	6	1.06	NA
5	5	0.70	0.62
6	16	1.03	0.43
7	11	0.60	0.43
8	6	0.69	NA
9	6	0.69	NA